

Bioinformatics for structural biologists

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Contents

- Predicting contacts from sequences using evolutionary covariance
- Bioinformatics for MR
 - Predicting domain structure
 - Informs phasing strategy, but also relevant earlier
 - Predicting tertiary structure
 - Homologous relationships identify search models for conventional MR
 - **AMPLE** does unconventional MR with *ab initio* models etc
 - Predicting quaternary structure
- Bioinformatics for structure-based function prediction
 - Using newer/less well-known data
 - Majoring on easily available servers/predictions

PREDICTION OF
STRUCTURE

PREDICTION (AND
EXPLANATION) OF
FUNCTION

```
>target_seq (s)
GYVELYRRSTIGNSLVDALDTLISDG
RIEASLAMRVLETFDKVVAETLKDNT
QSKLTVKGNLDTYGFCDDVWTFIVKN
CQVTVEDQSVISVDKLRIVACNSKKS
```

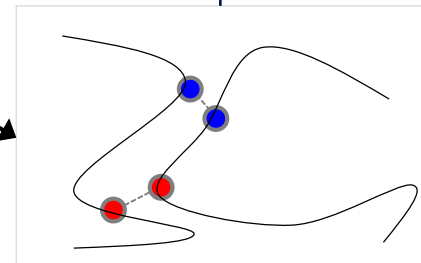
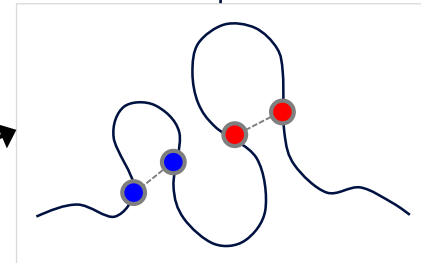
Multiple Sequence Alignment
of homologous sequences

...KDNTQSKLTVKGNLDTYGFC...
...KDNTQSKLTVKGNLDTYGFC...
...KNDANIAKLKLTFGDLDTY...
...RENTQSKLNVKGLLDTYGFC...
...KNENIVGKSKLIFKGLDTY...
...KNESNIAKSKLTFKGLDTY...
...KHESNIAKSKLTFKGLDTY...
...KNESGISKSKLTFKGLDHTY...
...KDNTQSKLTVKGLLDTYGFC...
...KSESGIAKSKLTFKGLDHTY...

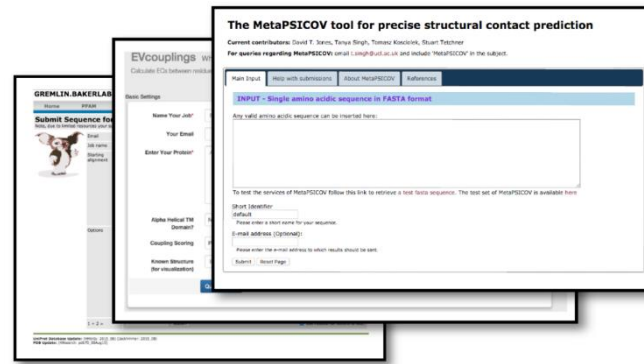
...KDNTQSKLTVK NLDTYGFC...
...KDNTQSKLTVK NLDTYGFC...
...KNDANIAKLKLT FKGLDTY...
...RENTQSKLNVK LLDTYGFC...
...KNENIVGKSKL FKGLDTY...
...KNESNIAKSKL FKGLDTY...
...KHESNIAKSKL FKGLDTY...
...KNESGISKSKL FKGLDHTY...
...KDNTQSKLTVK LLDTYGFC...
...KSESGIAKSKL FKGLDHTY...

Direct Coupling Analysis of
large MSAs plus Deep
Learning predicts contacts

Applications...



```
>seq
FASDGITF
DRSLFFGH
```

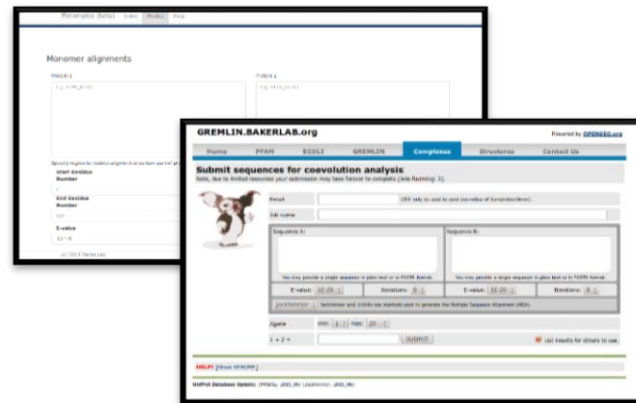


Intramolecular
contacts

Intermolecular
contacts if a
homooligomer

```
>seq1
FASDGITF
DRSLFFGH
```

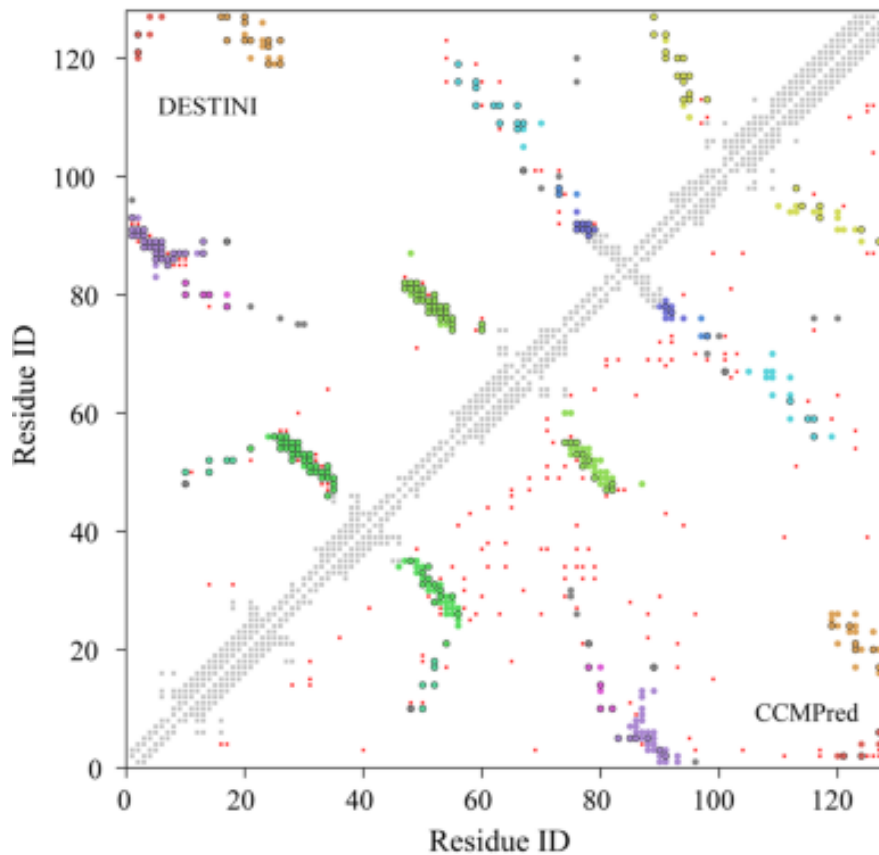
```
>seq2
NMHKLSDF
PLSERWAQ
```



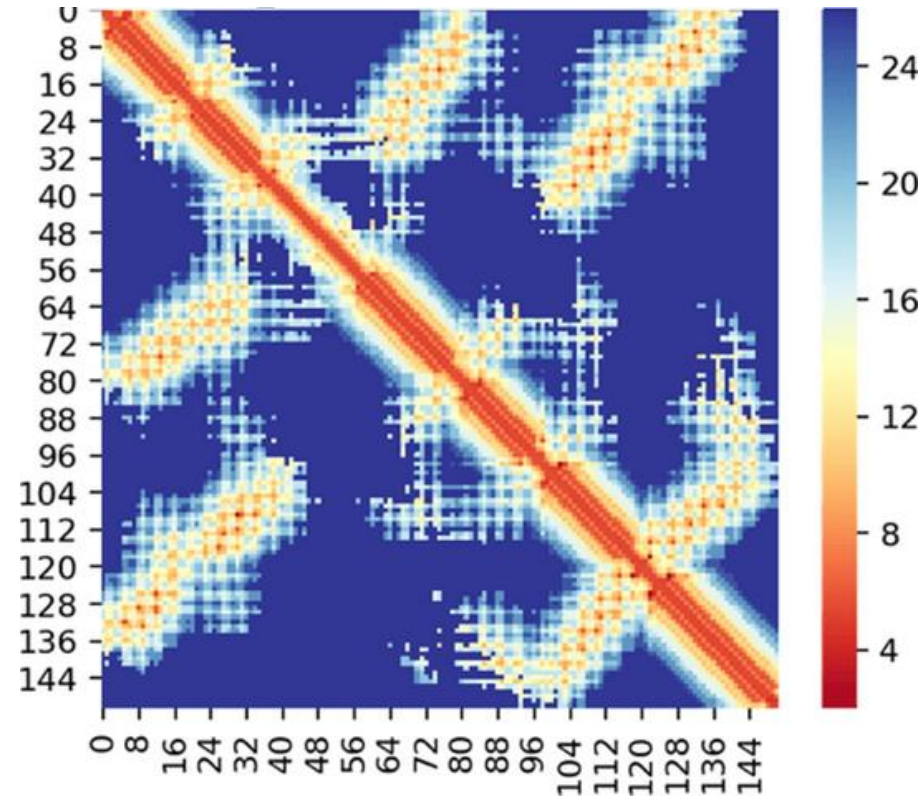
Intermolecular
contacts
[currently only
really reliable for
bacterial proteins in
operons]

ConKit can be used to visualise, analyse and convert contact predictions

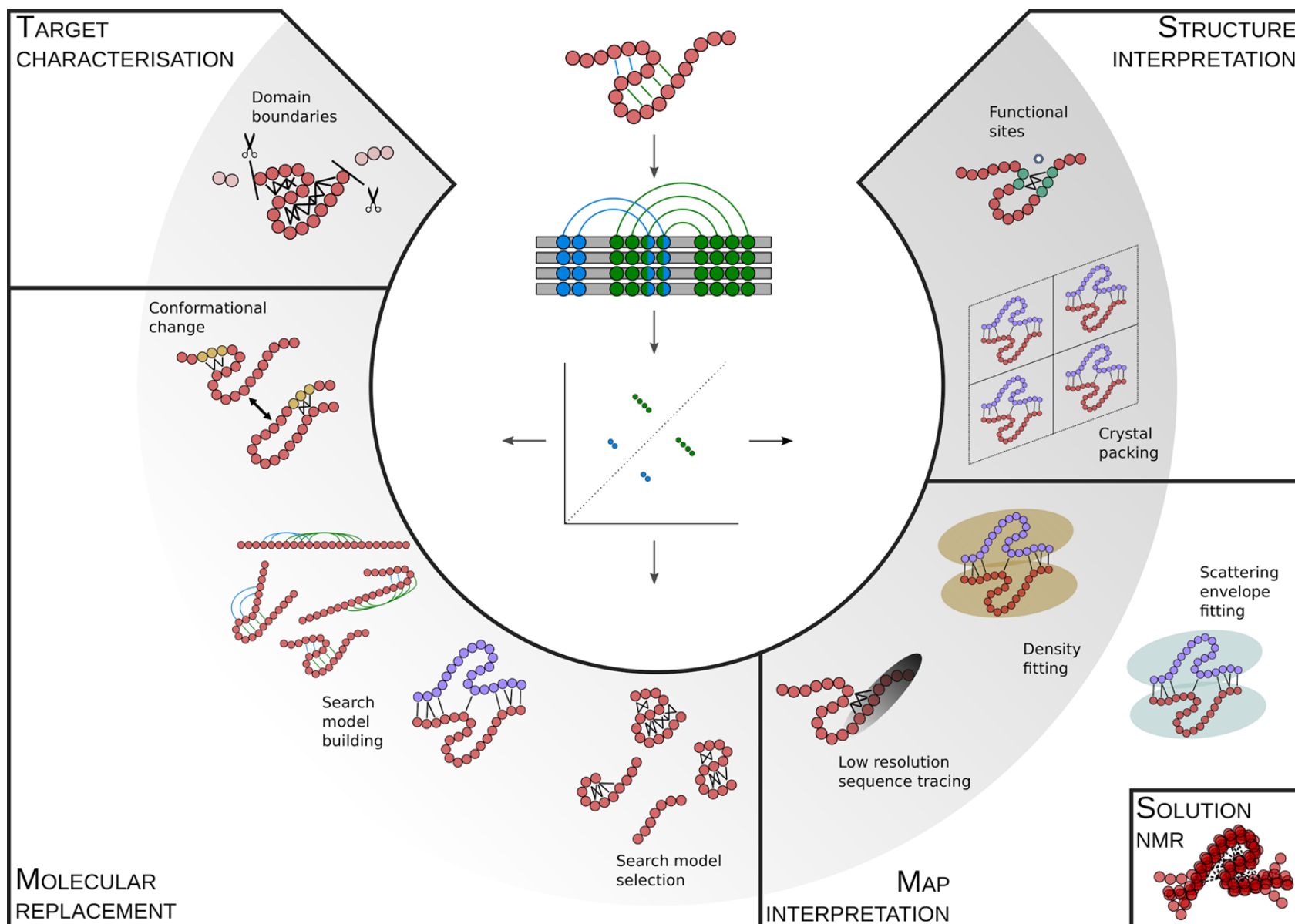
Simple predicted contact maps supplemented recently by distograms



Gao et al (2019). *Sci Rep* **9**, 3514



Adhikari (2020) *Sci Rep* **10**, 13374



Specific value of predicted contacts

- Structures unknown
 - Predicting domain boundaries (ConKit)
 - Making better *ab initio* models for MR, locally or at a server (AMPLE)
 - Work with new generation *ab initio* models from databases (MrParse; AMPLE)
- Structures known
 - Predicting how proteins interact (EVDock, Rosetta)
 - Validating the content of your crystal structure
 - (Predicting functional sites, highlighting conformational states)

Domains and tertiary structure

Which part to express for crystallisation?

Which parts of your crystallised protein might enable phasing by MR, or experimentally?

Recognising folded domains in your sequence

- How novel is your protein target? Recognising distant homology might make it less (or more!) interesting
- You might get extra ideas about its function to guide lab experiments, co-crystallisation, phasing (eg metal-binding sites)
- You might find the whole protein will not express/stay soluble/crystallise etc and want to deal with only part. You might well design construct to exclude disordered regions anyway.
- You might want to parse your protein into domains to explore different MR strategies

Recognising domains by homology with PDB, SCOP, CATH

- (PSI-)BLAST against the PDB might do it

sp|q5i8r5| (257 letters)

RID FRP1NV47014 (Expires on 03-31 21:44 pm)

Query ID gii74681646|sp|q5i8r5|
Description Trypsin-like serine
Molecule type amino acid
Query Length 257

Other reports: Search Summary

Graphic Summary

Show Conserved Domains

Query seq. 

Specific hits
Superfamilies
Multi-domains

Download GenPept Graphics

Chain A, Human Hepsin Protease In Complex With The Fab Fragment Of An Inhibitory Antibody
Sequence ID: [pdb|3T2N|A](#) Length: 372 Number of Matches: 1
[See 1 more title\(s\)](#)

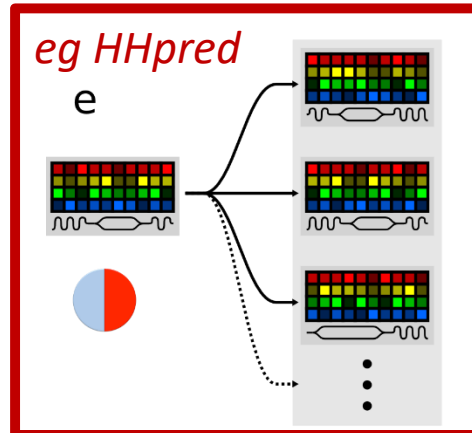
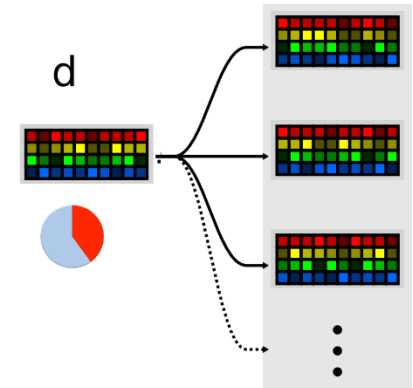
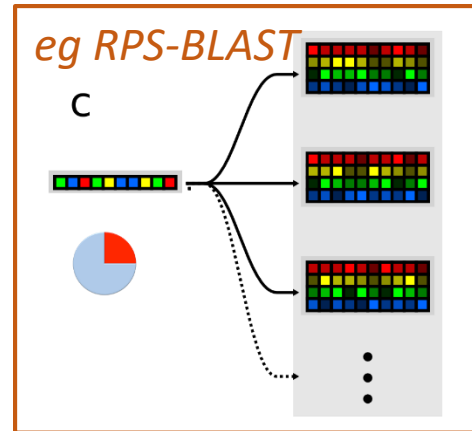
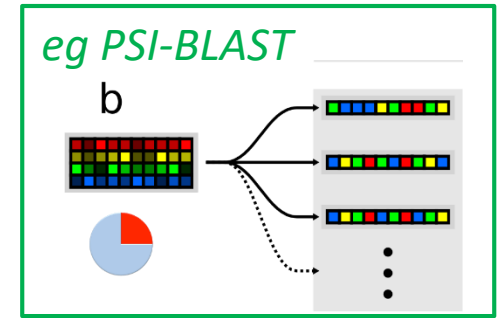
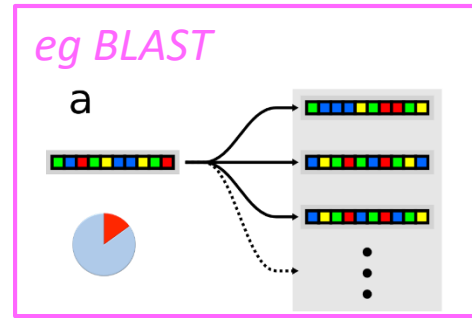
Range 1: 84 to 355 [GenPept](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Method	Identities	Positives	Gaps
142 bits(357)	6e-39	Compositional matrix adjust.	99/281(35%)	140/281(49%)	38/281(13%)
Query 1	MGLSTLSV-----LGLVTANYKLEKLPSGRIVGGYEVTFKFOYPLIASLEYGSHT	52			
+	LL ++SV L + + KLP RIVGG + T ++PW SL Y G+H				
Sbjct 84	QRLLEVISVCDPCRGRFLAAICQDCGRRKLPVDRIVGGRD-TSLGRWPQVSLRYDGAHL	142			
Query 53	CGGTLYNEKTIISAHC-----NIGSTSAWSASVHRHDLNEKAEEKSGSNHKII	101			
+	CGG+L + +++AAHC + + + AS H L +A G				
Sbjct 143	CGGSLLSGDWLTAACHFPERNRVLSRWRFAGAVAQASPHGLQLGVQAVVYHGGYL---	199			
Query 102	ERISHPOYDLN-DDSSNDVSVWKIAAPGNKT---SGIVLDSGKVSSDGTLLKVIWGTT	157			
+	P D N +++SND+++ ++P T + L + + DG + V GWG T				
Sbjct 200	-----PFRDPNSEENSNDIALVHLSSPLPLTEYIQPVCLPAAGQALVDGKICTVTGWGNT	254			
Query 158	TSGGDVSKVLLLEVKVPVFNIDKCKKA--YSTLDTASQFCAGYPEGGKDSCQGDSSGGPIFI	215			
+	G + VL E +VP+ + D C A Y FCAGYPEGG D+CQGDSSGGP				
Sbjct 255	QYYGQAGVLQEARVPIISNDVCNGADFYGNQIKPKMFCAGYPEGGIDACQGDSSGGPFVC	314			
Query 216	EEKGVAT----LVGVVSWGRGCALKGYPGVYTRVSKVLDFI	252			
+	E+ T L G+VSWG GCAL PGVYT+VS ++I				
Sbjct 315	EDSISRTPRWRLCGIVSWGTCALAQKPGVYTKVSDFREWI	355			

Recognising domains by homology with PDB, SCOP, CATH

- Harder cases require a more sensitive tool. I recommend HHpred. Used by **MrBUMP** to find homologues to use as search models



- HHPRED works by comparing HMMs of **alignments**, not just single sequences
- Matching of **secondary structure** also scored

```
Q ss_pred          EEEEECCCCCEEECCCEEECCCCCEEECCCCCEEEEEEEEEECCCCCEECCCCCEEECCCCCEEECCCCCEEEEEEEECCECCC
Q ss_conf          69998345548954135307841599962465599841984089872056055223874167786189731880346774
Q gi|60495103|em   250 GLFIFLESKNITLSRLQMHYMHGLGIVSQYTENITMDRVKCAPRPDSGRLLAAASADMHHFSGKGKVIIDSCYFAGAQDDP    329 (605)
Q Consensus        250 ~-~--nV-ie-iti-s-gi~~~~~niti-i~~~~r-s-aDg~~~~i-nC-f~-DD~    329 (605)
                    ++.+.|+|+++++++.|+++.|.l++++.++.+.+.+.      +||+++|.++ ++|+|+|. .|+||
T Consensus        333 ~I~~~~ni-I~~iti-s~~~~i~~~~s~~~~in~~~~-----n-DGI~i~s~n-v-I~N~i~--gDd~    404 (608)
T 2uvf_A           333 LMTLRGVENVYLAFGTVRNPAPFHGIMLENHNHVAVANGLTHQTYPDAN---- -NGDGIEFGNSQN-VVMFNFFD-TGDDC    404 (608)
T ss_dssp          SEEEESEEEEEESCEECSSCSSEESSCEEEESCCEETCT-----TCSSEEEESCCE-EEESCCEE-CSSCS
T ss_pred          EEEEECCCCCEEECCCCCEEECCCCCEEEEEEEEEECCCCCE-----CCCCCEEECE-EEESCCE-CCCC
T ss_conf          7999605773899889972478617871444389801798288876-----6444178734772-799524520-58764
```

HHpred tips and warnings

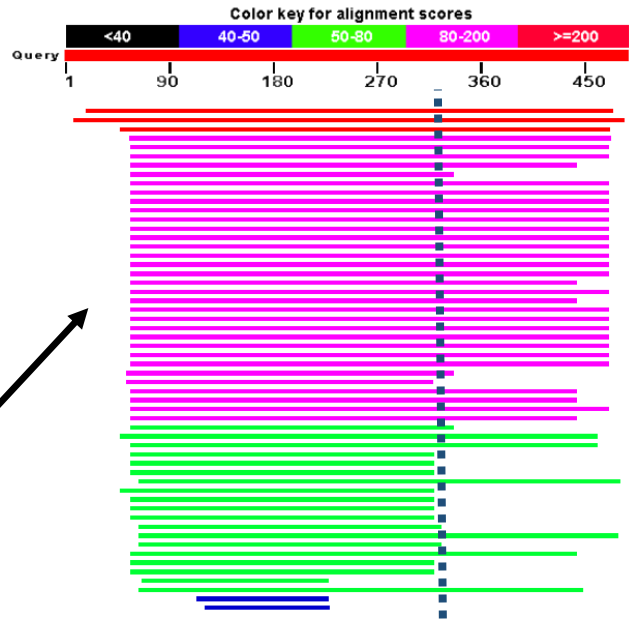
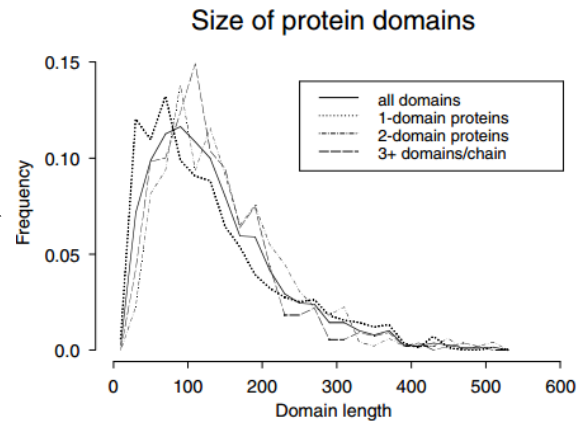
- **Probability** (0-100) is generally a good guide....
- ...but statistics can mislead for **unusual** protein sequences eg coiled-coil, low-complexity, Cys-rich
- Consider if the match makes biological **sense**!
- Look at the matched region
 - Is it a complete domain/structure?
 - If partial, could it reasonably fold?
- Can make reasonable homology models too
- If your query contains multiple domains, '**zooming in**' on particular regions can improve scores and show results not previously seen since
 - Score contains an element favouring similar lengths
 - Only 250 results are shown: can easily get this number for a common domain, making other results invisible!
- **MrBUMP** can search for MR search models with HHpred, most easily at CCP4 online or CCP4Cloud

Recognising matches to domains in **sequence** databases, Pfam, Smart etc

- HHpred is also an excellent, sensitive way to search against these. The same rules for interpretation apply. Matches, even distant ones, can obviously shed light on function
- However, for **structural** purposes need to remember that
 - Many Pfam families don't have structures so that domain limits are much less precise.
 - Many examples where a structure redefines previous Pfam sequence-only domain boundaries
 - Large Pfam entries especially DUFs often turn out to have multiple structural domains
 - Pfam entries for repeats sometimes contain multiple copies

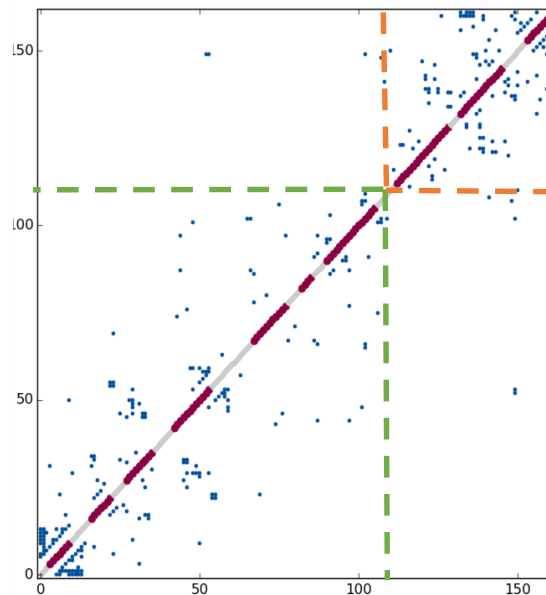
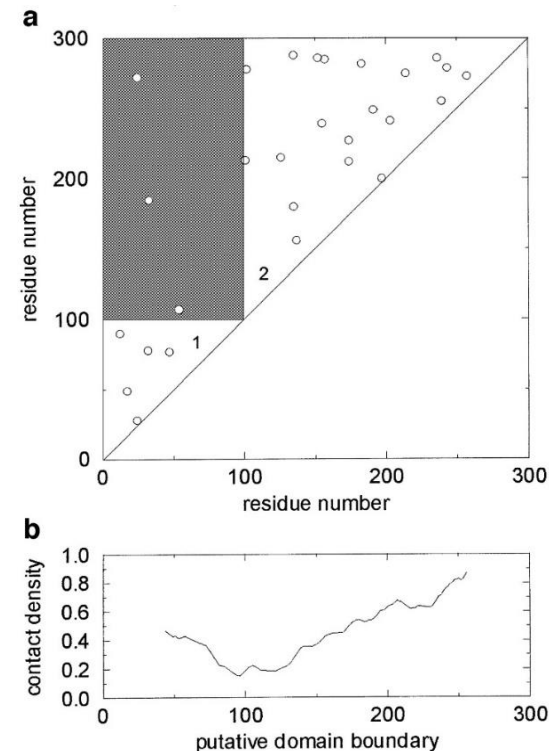
And if the domains
can't be matched?

- *ab initio* approaches to domain boundary identification
 - BLAST matches in sequence databases. Domains are often found in different combinations
 - Secondary structure pattern. PSI-PRED or Jpred4 (faster server)
 - Domain guess by size (server defunct)
 - Contact predictions (ConKit)

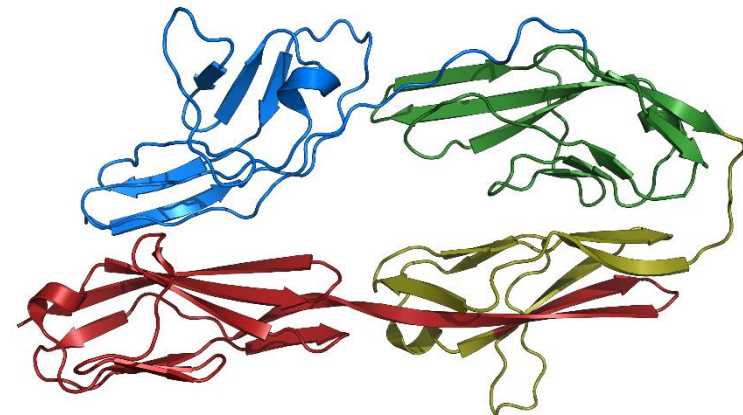
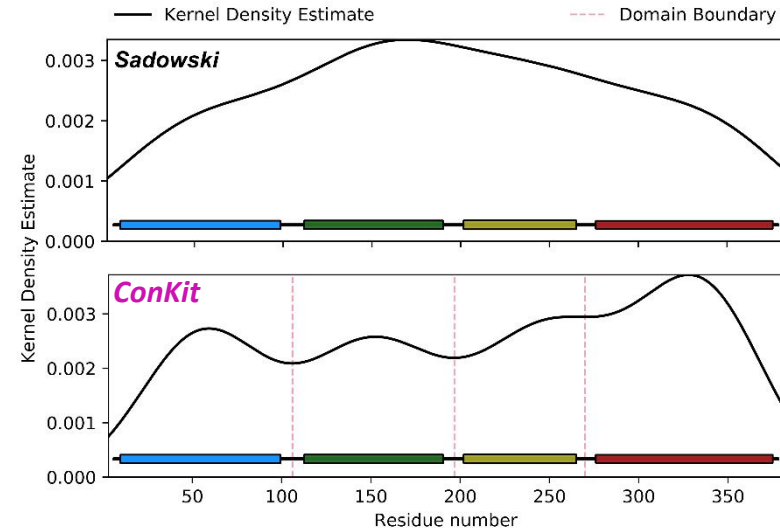
[illegible]

Predicted contacts for defining domains

- Domain boundaries required for individual expression *in vitro* and helpful for fold recognition and modelling *in silico*

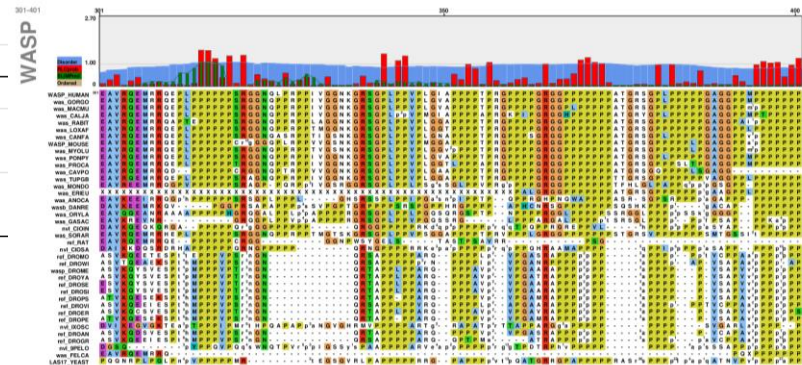
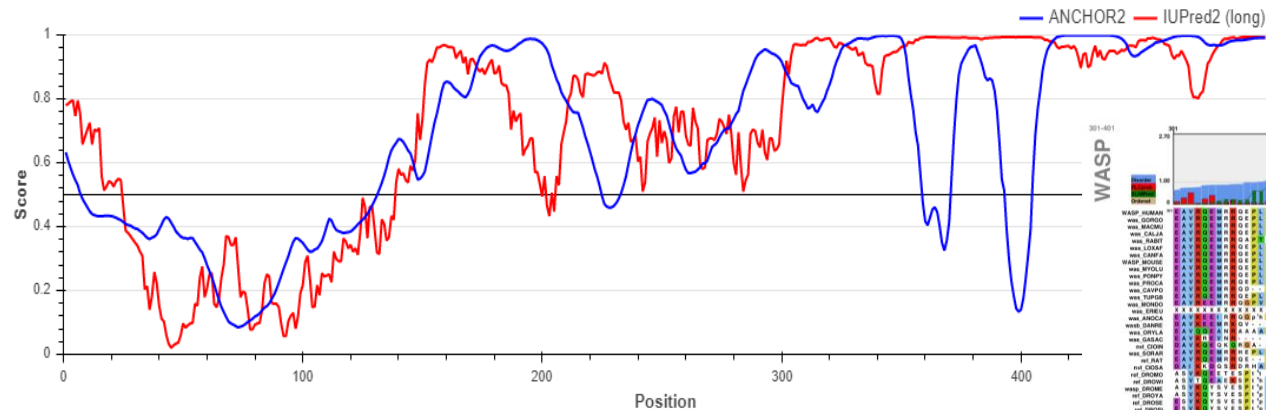


DUF4158 – two helical domains?



Intrinsic disorder prediction

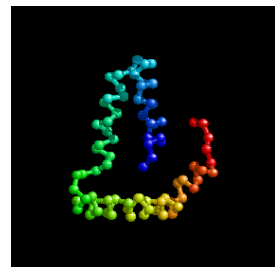
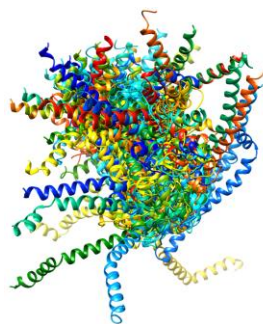
- Not all proteins and protein regions fold into stable structured domains. ID proteins and regions will not crystallise (alone)
 - There are many predictors, all performing roughly equally well
 - I recommend IUPred (fast) and MetaDisorder (slow but good)
 - Can also look for short interaction motifs in ID regions (ANCHOR, SlimPred)
- 



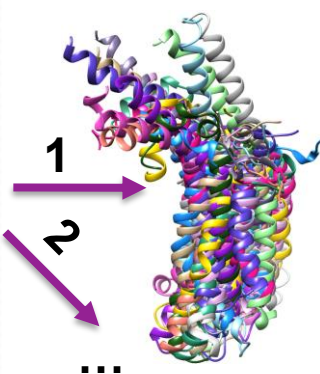
ab initio models and MR

Building **ensemble** search models with **AMPLE**

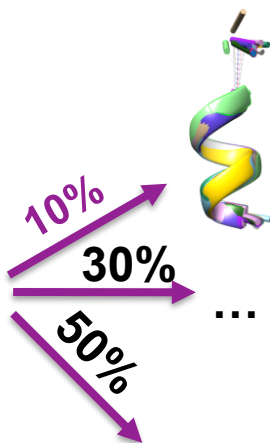
ab initio
models
(ROSETTA/
QUARK)



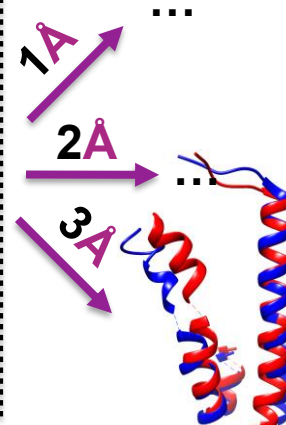
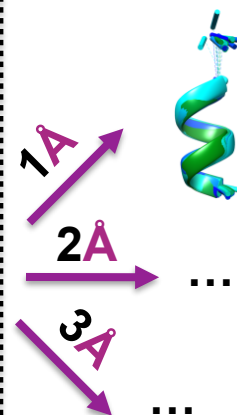
cluster
(SPICKER)



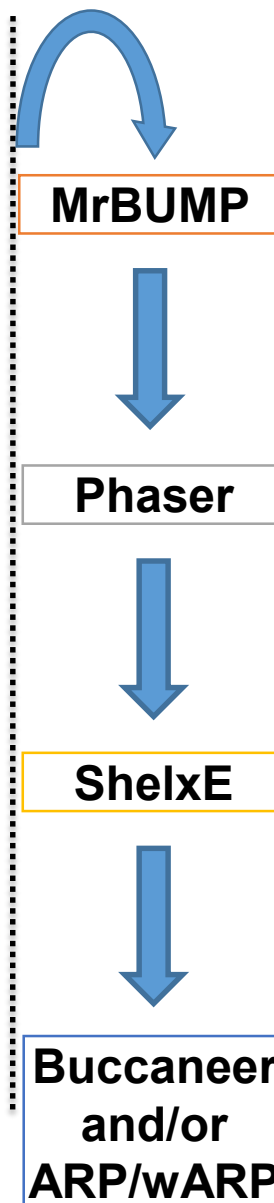
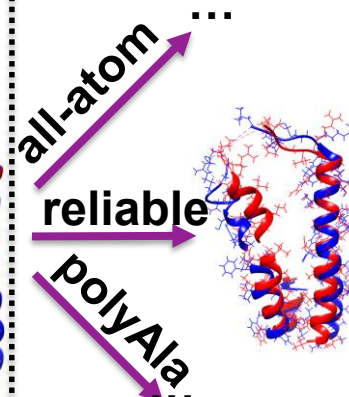
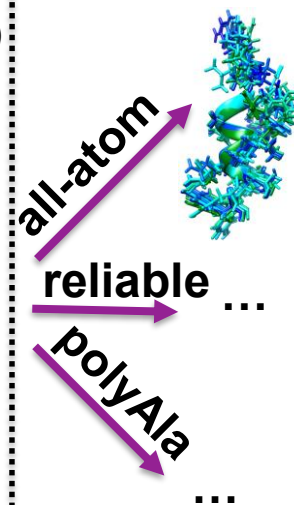
truncate
(THESEUS)



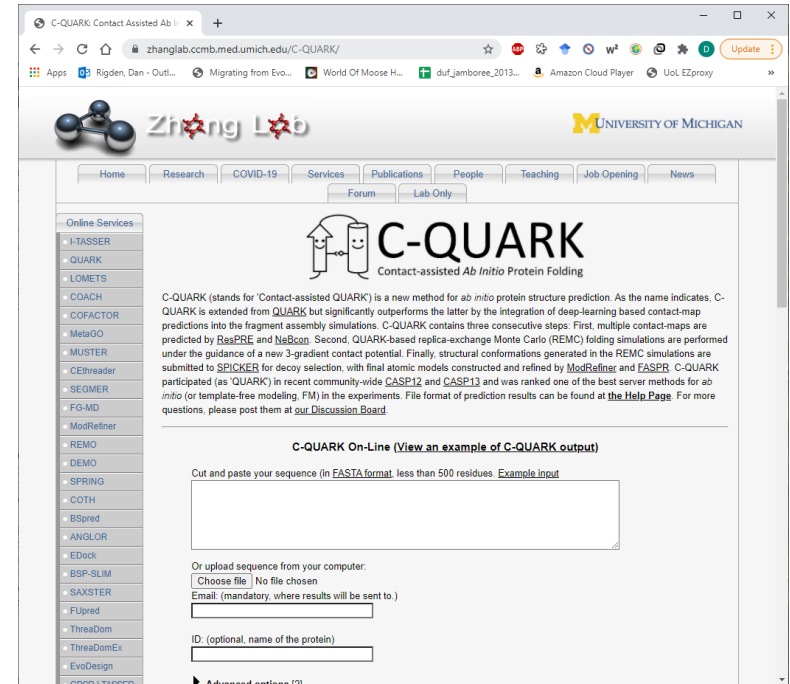
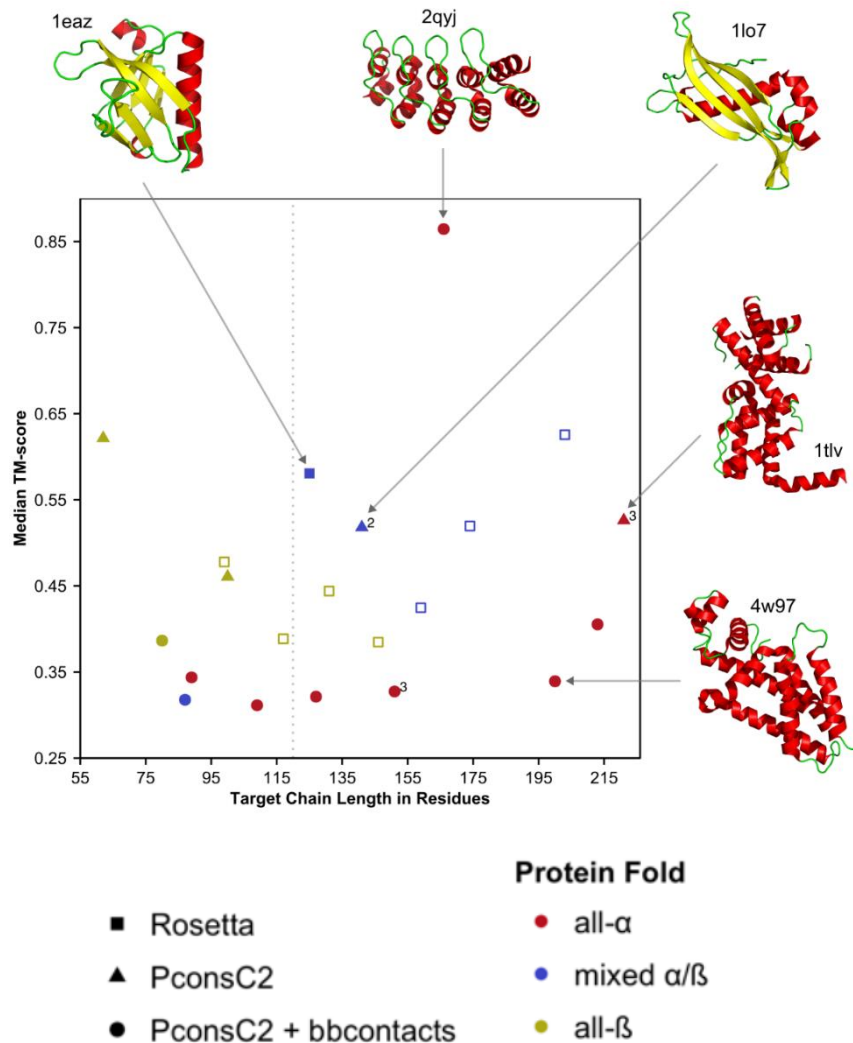
cluster
(MAXCLUSTER)



side chains



Contact predictions improve local and server *ab initio* models

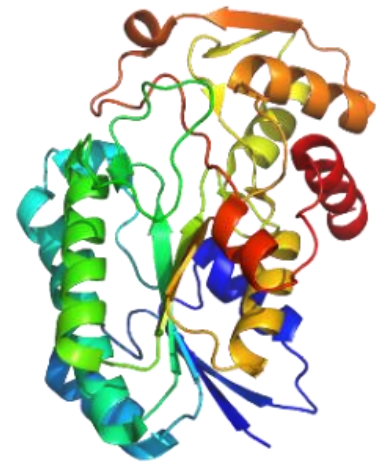
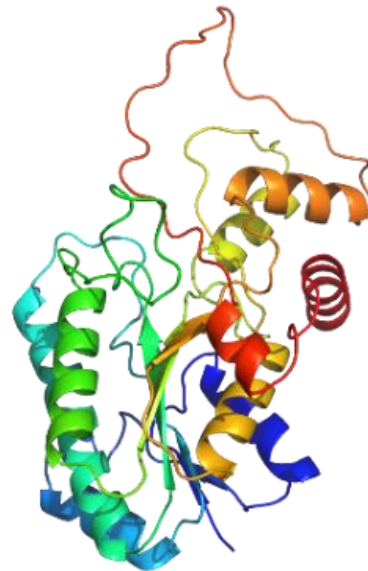
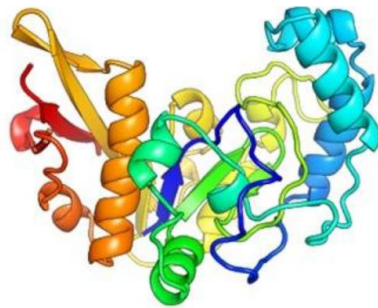
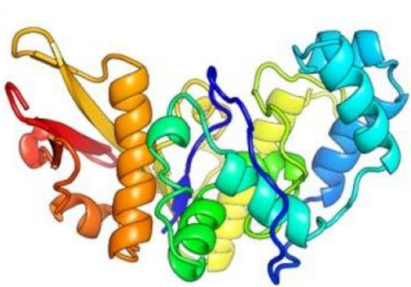


models.tar.gz

i2, Cloud, CCP4 online,
command line

Using *ab initio*, contact-driven models from databases

- GREMLIN metagenome repository
- Models for 614 Pfam families (predicted high quality)
- 30 best models per family, or single best
- Scored according to predicted reliability
- gremlin2.bakerlab.org/meta.php
- Pfam and InterPro plan to integrate models made this way
- PConsFam database
- Models for 13,617 Pfam families
- Single best model per family
- Scored according to predicted reliability
- pconsfam.bioinfo.se



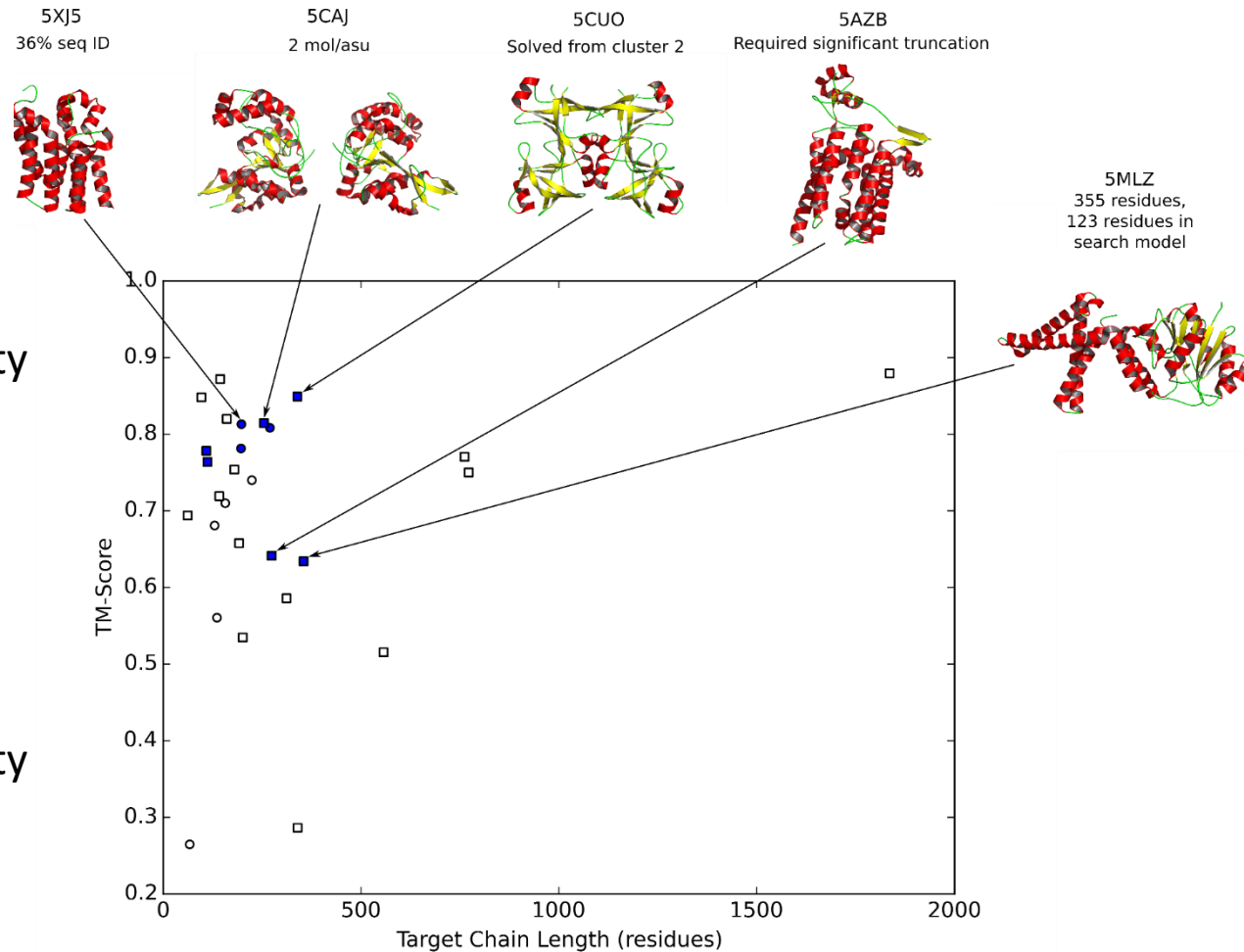
AMPLE results with GREMLIN database deposits

- Targets

- 1-12 mol/asu,
- 1.5-3 Å resolution,
- 62-1836 residues,
- 36-100% sequence identity to modelled protein

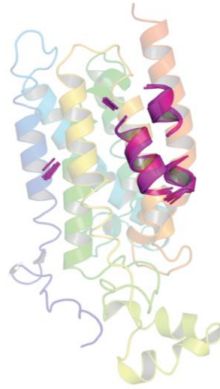
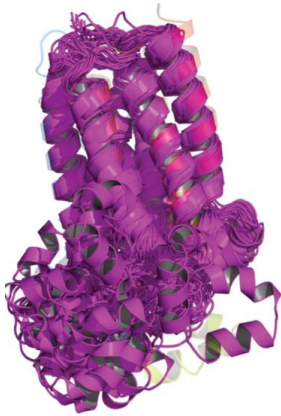
- 9/27 succeed

- 1-3 mol/asu,
- 1.5-2.9 Å resolution,
- 109-355 residues,
- 36-100% sequence identity to modelled protein)

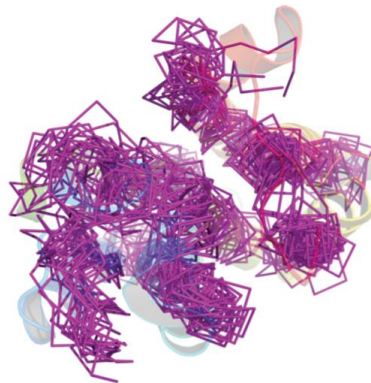


AMPLE working on *ab initio* model database deposits

GREMLIN



PCONSFAM



MrParse collects info relevant to MR

Diffraction information e.g. pathologies from Phaser

Protein features relevant to MR strategy choice:
secondary structure (JPred3), TM helices (Topcons) and coiled-coil (Deepcoil)

An additional Table highlighting models available in databases will be coming soon

MrParse Homolog Search

Version: 0.1.8

This is the alpha-release of MrParse: a program to find and analyse search models for Crystallographic Molecular Replacement. The program is being developed by [Dan Rigden's group](#) at the University of Liverpool

For more information about MrParse, please

MrParse is currently under development and

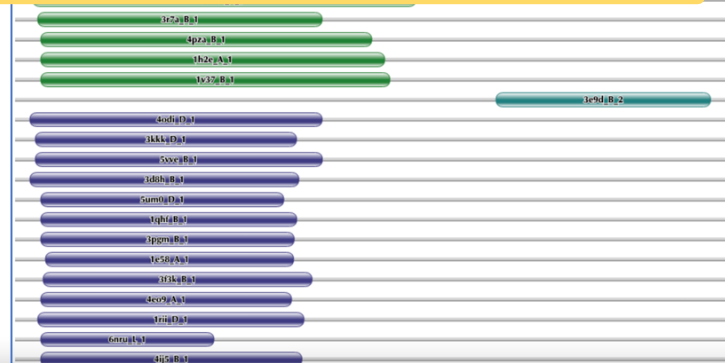
name	Resolution	Space Group	Has NO
3dcv-sf	1.75	P212121	false

Homologs	
Name	
3dcy_A	<i>in silico</i> prediction
4emb_D	
3gp3_D	
3e9d_B	
3r7a_B	
4pza_B	
1h2e_A	Model Identifier
	PF00300.19

in silico predictions

Model Identifier	Coordinates url	Provider	Created	Seq. Ident.	Coverage	Qmean Average Local Score
PF00300.19	PF00300.19	PconsFam	16-Jan-2017	100	70	-6.6

lv37_B_1	lv37	140	3	10-144	133	112.6	14032.01	0.991	0.37
3e9d_B_2	3e9d	2.00	4	186-268	81	49.4	9521.01	1.021	0.35
4odl_D_1	4odl	2.60	5	6-118	111	82.6	12447.07	1.021	0.35
3kkk_D_1	3kkk	2.08	5	8-108	99	81.6	11524.19	0.976	0.38
5vve_B_1	5vve	1.70	5	8-118	109	91.5	12742.52	1.006	0.36
3dRh_B_1	3dRh	2.01	5	6-109	102	67.7	11881.49	1.053	0.33
Sum0_D_1	Sum0	1.85	5	10-103	92	71	10779.23	0.976	0.38
1qhf_B_1	1qhf	1.70	5	10-108	97	67.2	11391.97	1.021	0.35
3pgm_B_1	3pgm	2.80	5	10-107	96	64.6	11158.65	1.021	0.35
1e58_A_1	1e58	1.25	5	12-107	94	56.4	10980.34	1.053	0.33
3f3k_B_1	3f3k	1.75	5	11-114	102	81.4	12868.58	1.102	0.3
4eo9_A_1	4eo9	2.45	5	10-106	95	53.3	11285.86	1.085	0.31
1rl1_D_1	1rl1	1.70	5	9-111	101	48.6	11665.18	1.136	0.28
6nuv_L_1	6nuv	2.50	5	10-76	65	46.2	7353.57	1.069	0.32
4j5_B_1	4j5	1.50	5	10-110	99	58.9	11084.74	1.053	0.33
3r7a_B_2	3r7a	1.84	6	183-213	29	9.85572	3012.76	1.085	0.31
4odl_D_2	4odl	2.60	6	195-212	16	2.59048	1945.35	0.932	0.41
3kkk_D_2	3kkk	2.08	6	196-212	15	1.93261	1806.19	0.991	0.37
1h2e_A_2	1h2e	1.69	6	196-214	17	2.49718	2028.55	0.976	0.38
5vve_B_2	5vve	1.70	6	196-214	17	2.60219	2097.49	0.976	0.38



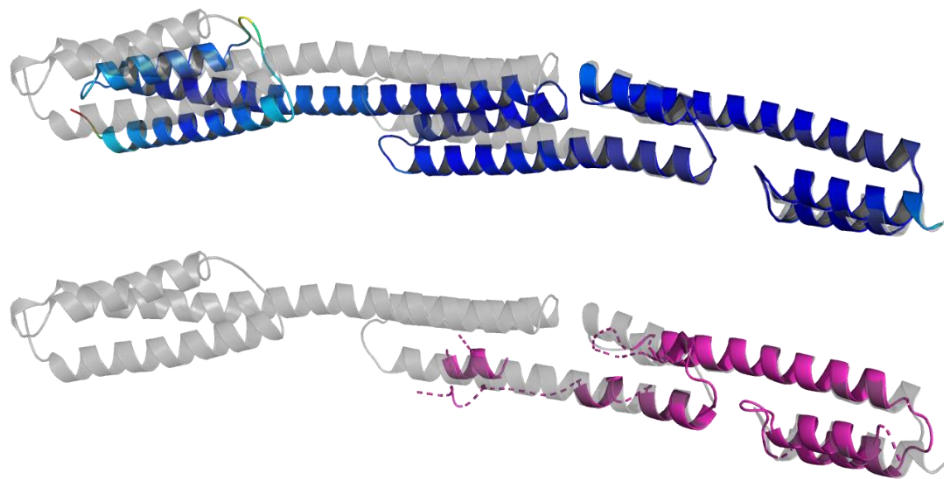
Sortable table of homologs found by phmmer (or HHpred in the future). Ranked by eLLG by default

Visual representation of matches,
colour-coded by MrBump 'domain'

AMPLE working on AlphaFold2 results

- The very best models may not need ensembling
- 24/30 worked straightforwardly
- A further 3 needed AMPLE truncation
- AMPLE single structure mode can
 - Truncate models according to the estimated error in the B-factor column
 - Convert the estimated Å error to a B-factor (borrowing from Randy Read's script)

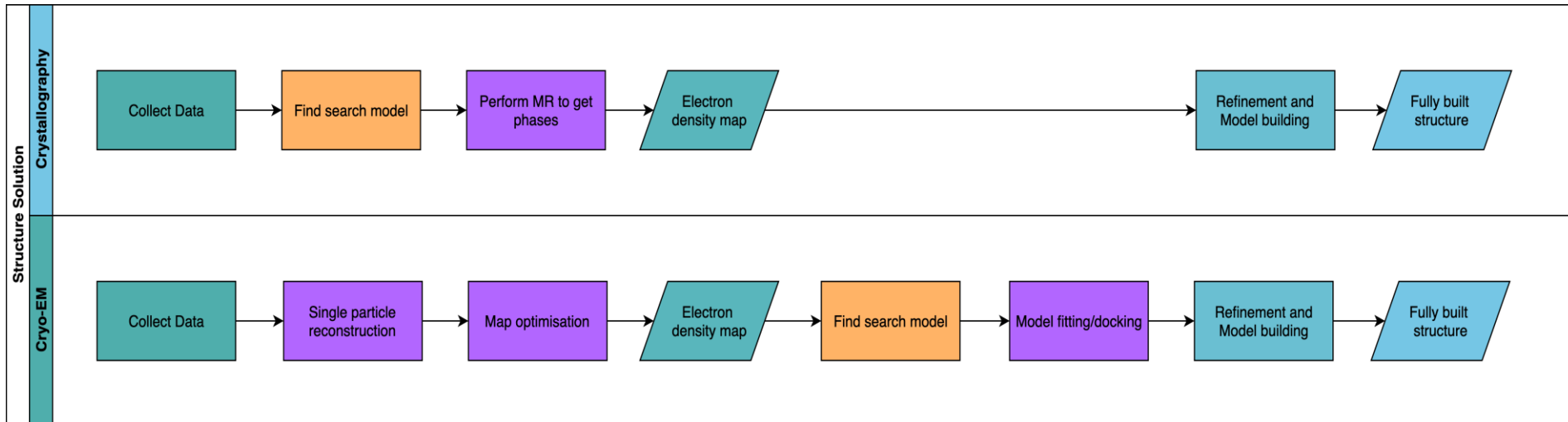
```
ample -single_model <PDBFILE>  
-truncation_method bfactors /  
-truncation_method errors
```



T1030. 3.03 Å

Solves with 18-44%
truncated versions

... and not forgetting Cryo-EM



- **MrBUMP** repurposed to find and place structures in Cryo-EM maps
- In the future will be able to place in silico predictions found with **MrParse** too

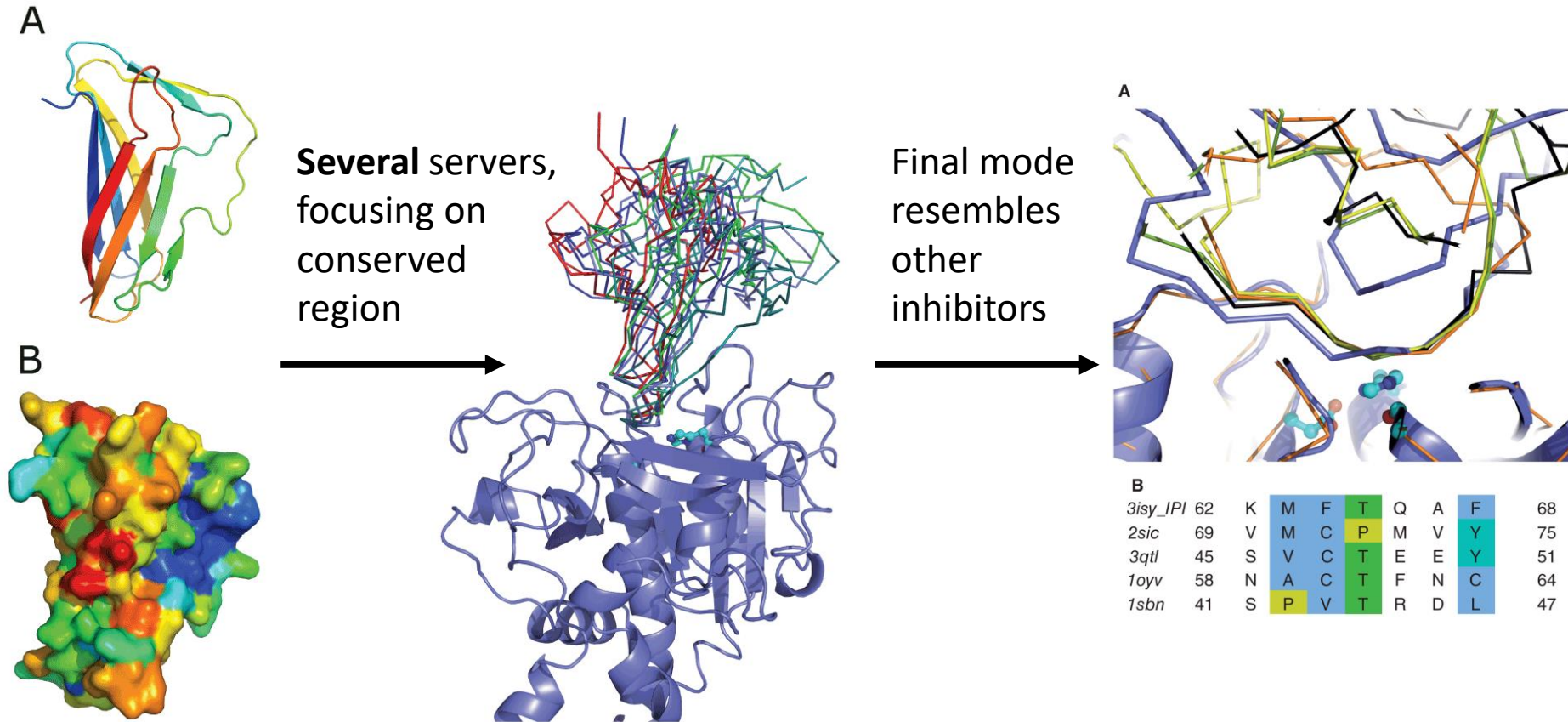
Quaternary structure

Intermolecular interactions

Predicting protein-protein interactions

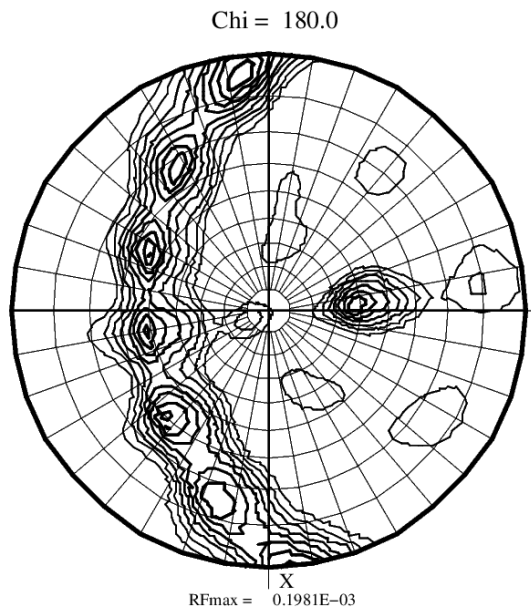
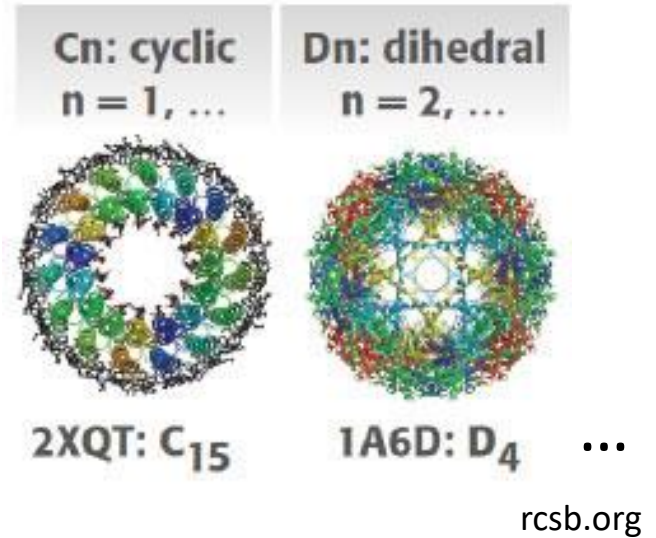
- Relevant to MR eg proteins A and B are cocrystallised but neither alone solves. An accurately predicted complex, being larger, might solve
- Many methods predict complexes based on steric complementarity plus other scoring functions
- Recommendable servers include
 - ClusPro, the best performing docking method
 - Haddock, which has a good server with different modes
 - Each allows inclusion of other information eg predicted interface residues
 - Symmetric docking at ROSIE server

Multiple methods in bioinformatics: *B. subtilis* IPI docking to protease

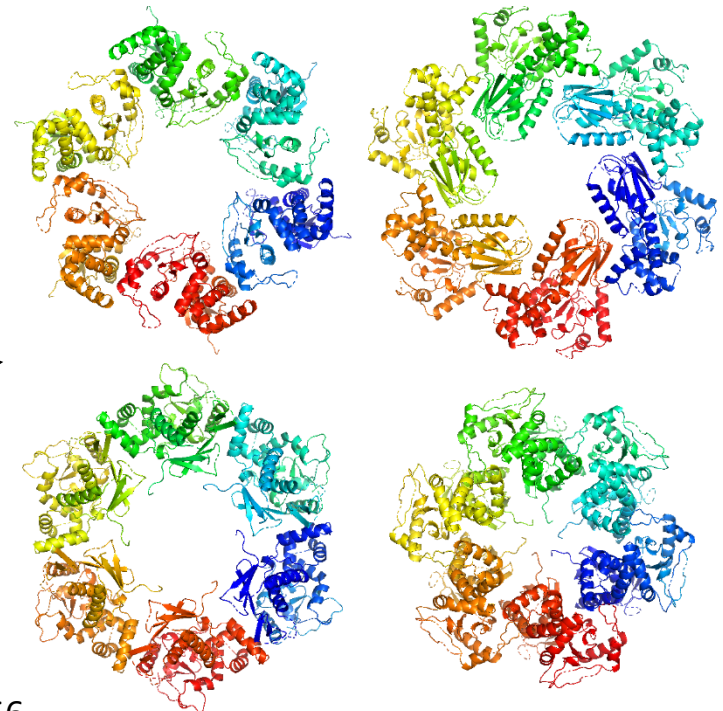


ROSIE symmetric docking for oligomers

- Only cyclic (C_n) or dihedral (D_n) symmetry at server
- Clues from self rotation function may be available

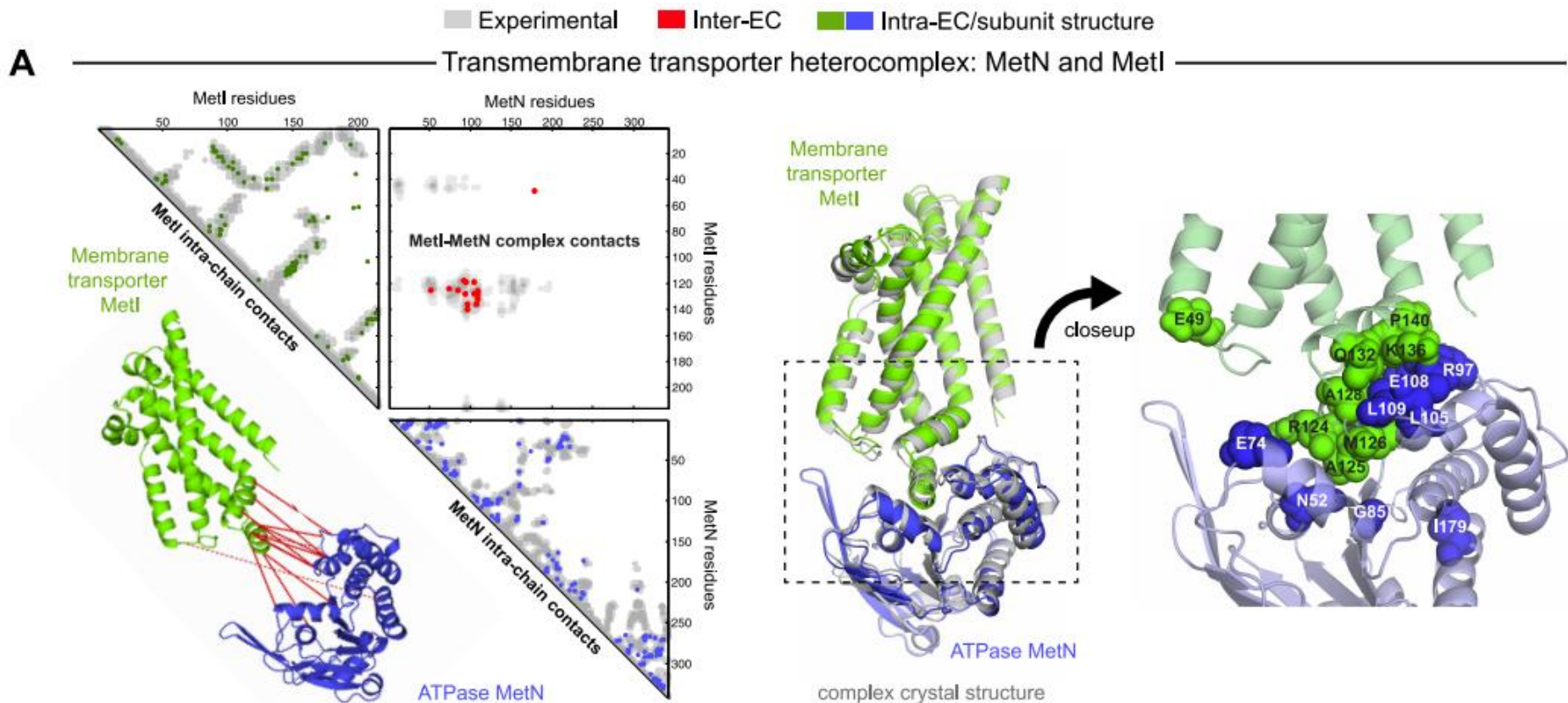


Generate
hexamers

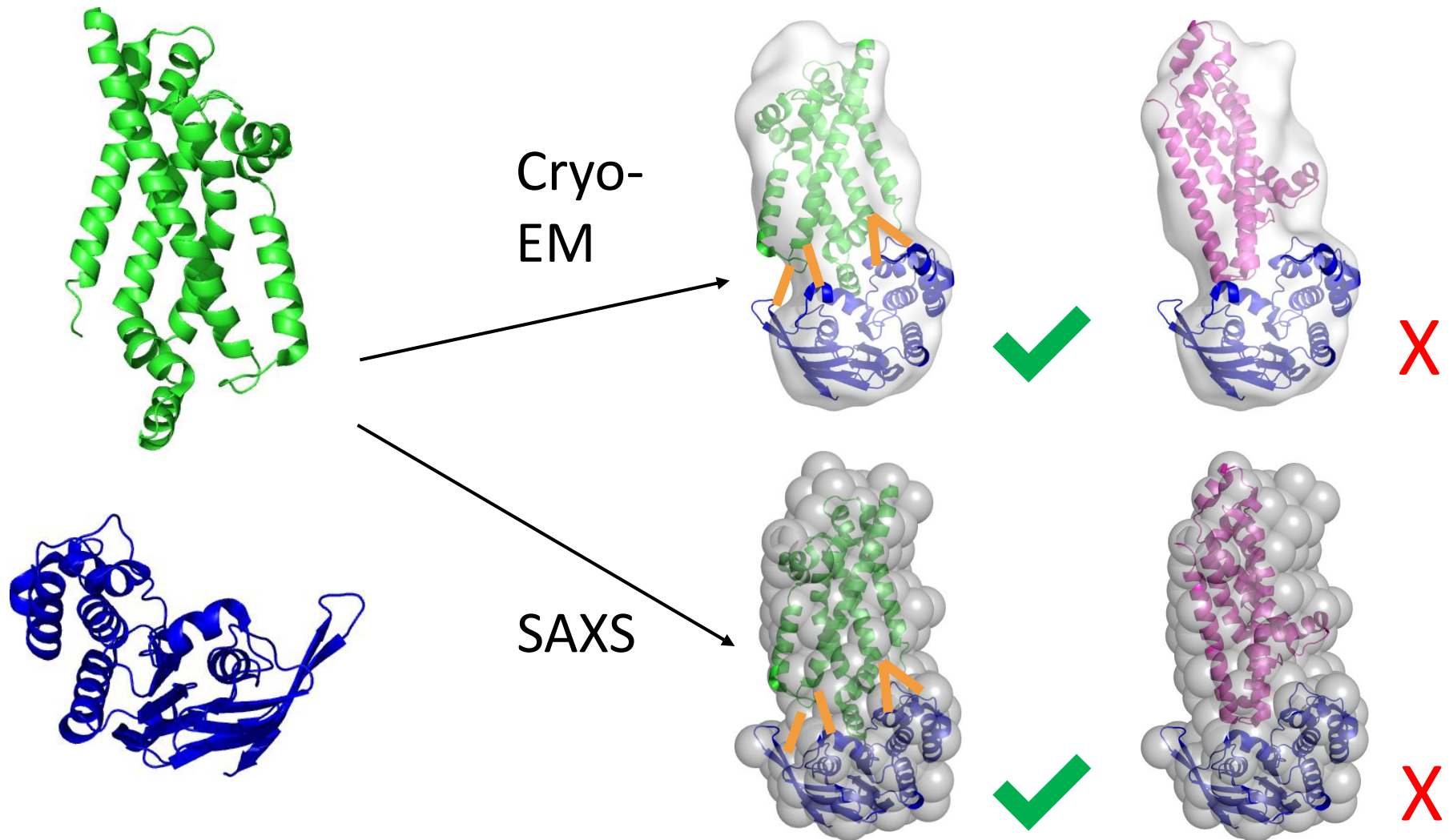


Andre et al. (2009) PNAS 104:17656

Using predicted contacts to help predict complexes



Fitting to low-resolution envelopes



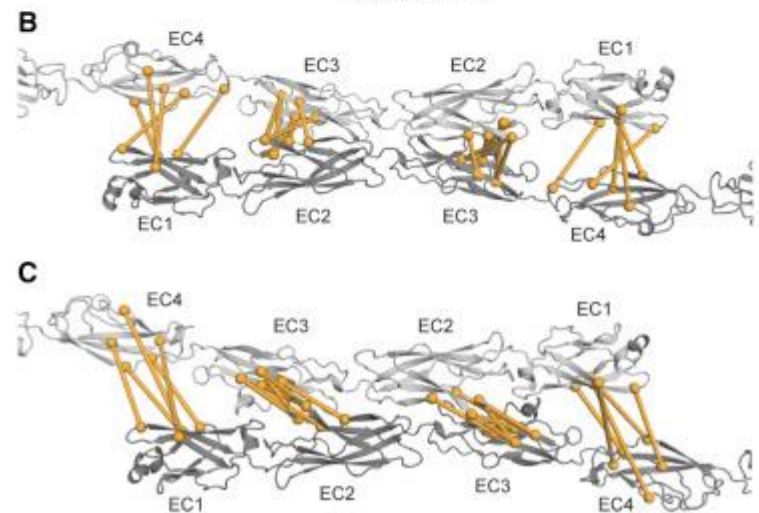
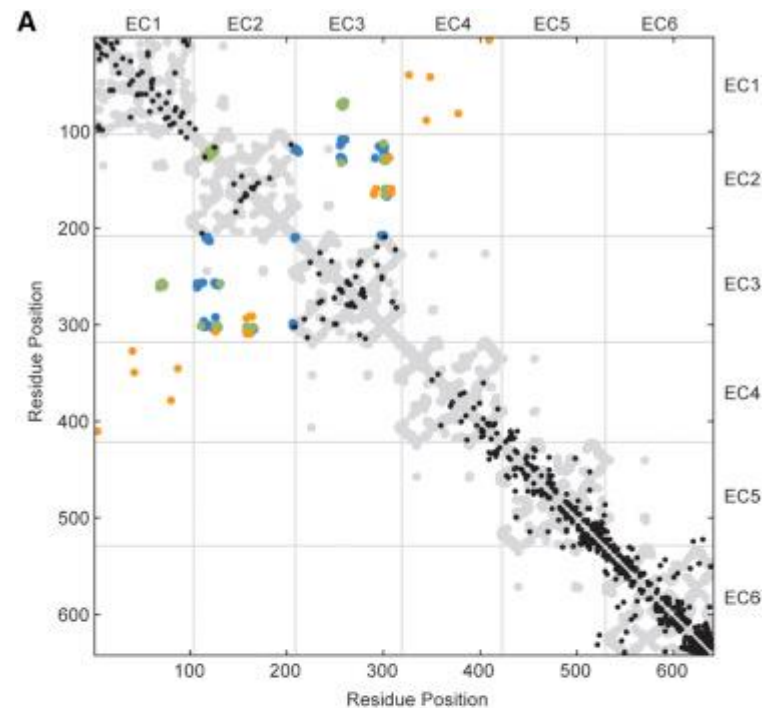
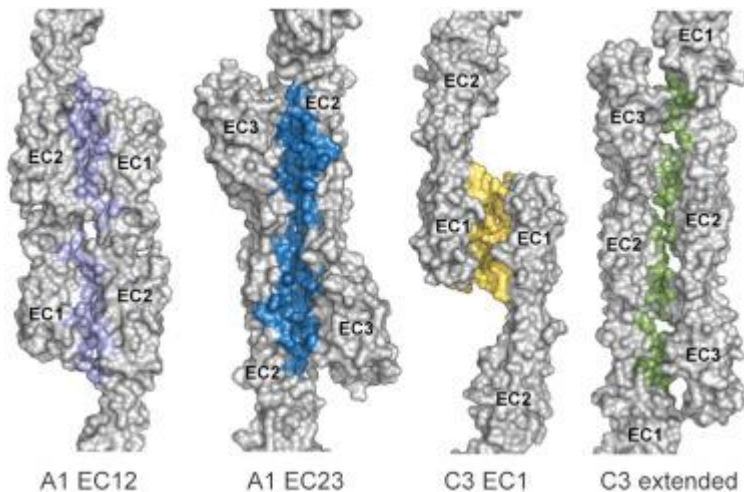
Structure-based function interpretation

What is the biologically relevant quaternary structure?

Where are the functional/catalytic sites?

Validating crystal structure contents

- **PISA** is an excellent general method, but contact predictions help in some cases
- Crystal showed various ways in which protocadherins could interact
- Predicted contacts supported only some of the modes
- **PISA-cov** is on the way...

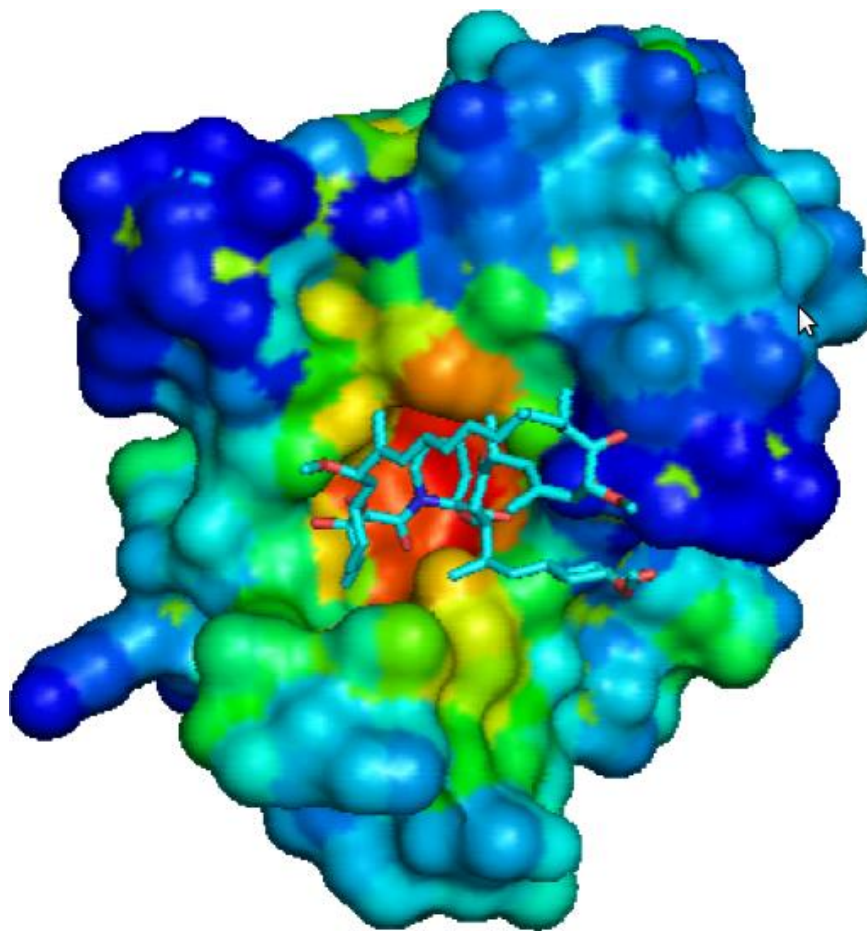


Some lesser-known structure-based function annotation methods

- Finding functional sites is based on their being different somehow to the rest of the protein surface. Important generic methods are based on
 - Shape (castP, ProFunc, PyMOL)
 - Electrostatics (PyMOL, APBS)
 - Evolutionary conservation (Consurf)
- Less well-known but valuable characteristics are
 - Statistics of surface atom 'triangles' (STP)
 - Probe interaction energetics (ISMBLab)
 - Predicted pKa values (THEMATICS/POOL)

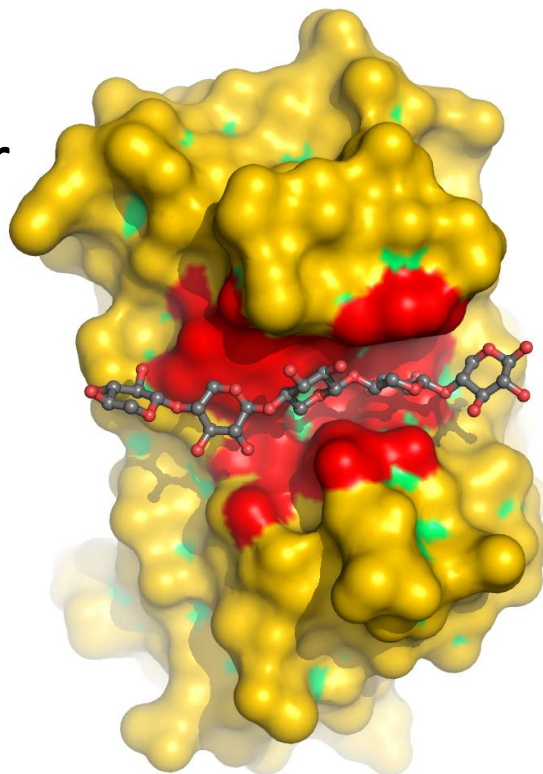
Binding sites from statistics

- STP (surface triplet propensities)
- 13 atom types $\rightarrow$ 455 triplets
- Distribution in binding vs non-binding sites varies
- Designed for small molecules, works on PPIs, including flat surfaces

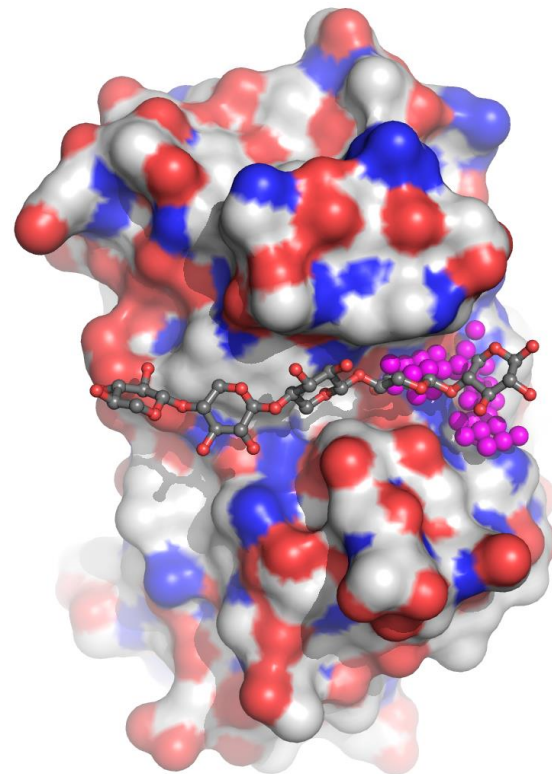


Different probes for different binding sites

- Hydroxyl group can be used to probe for carbohydrate binding sites
- Phosphate oxygen used for binding sites of phosphorylated ligands



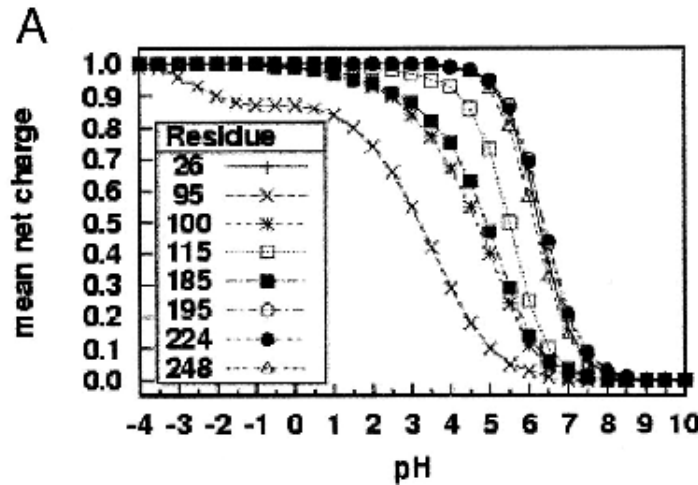
ISMBLab



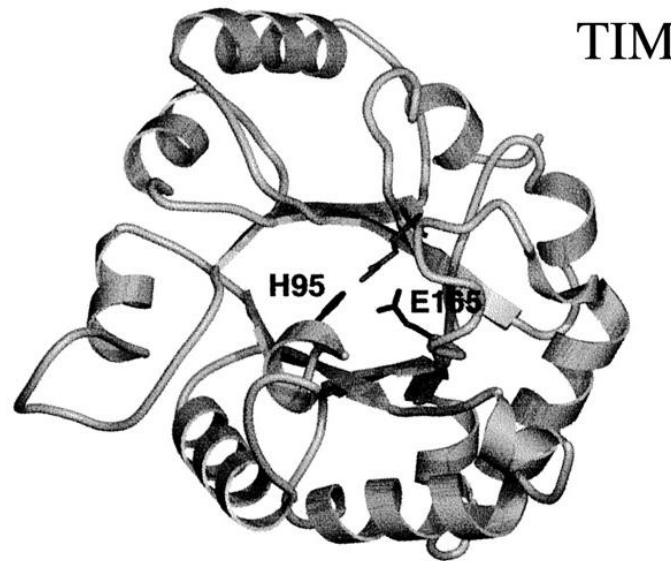
SiteHound

Theoretical microscopic titration

- Computer analysis of a reliable protein structure can predict pKa values for acids and bases. Residues with perturbed pKa values are possible catalytic residues, especially if clustered.



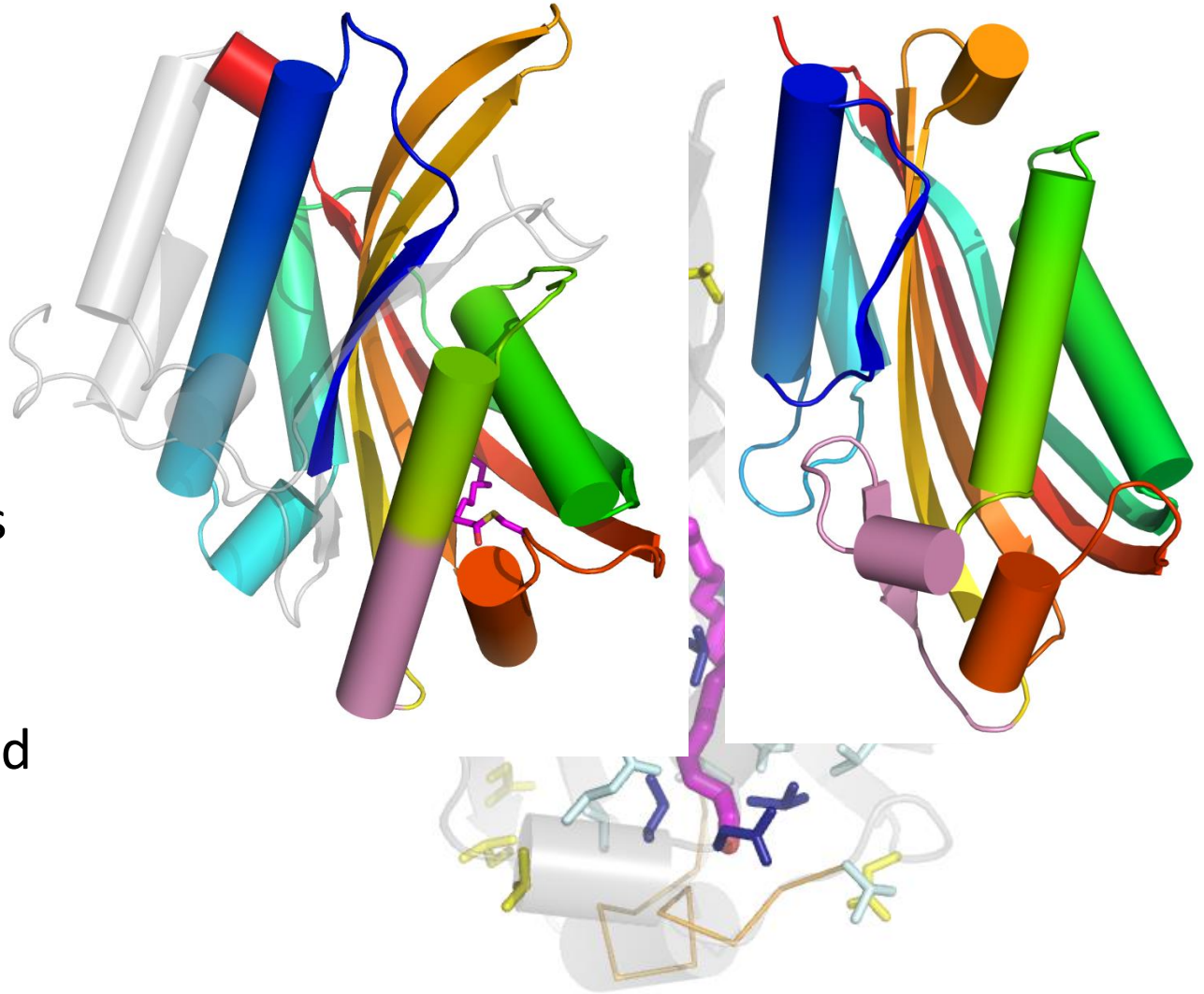
pKa of His95 is atypical compared to other His residues in enzyme



His95 and other residues with atypical pKa cluster at catalytic site

Multiple methods in bioinformatics: Structure comparisons of Evf

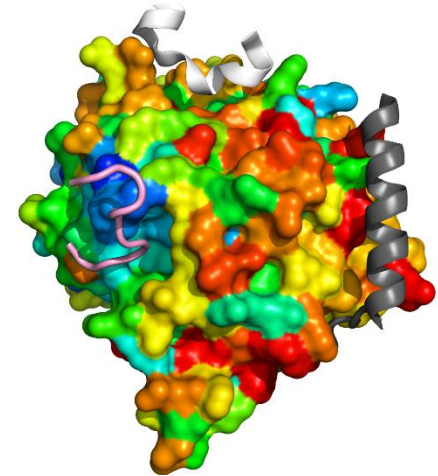
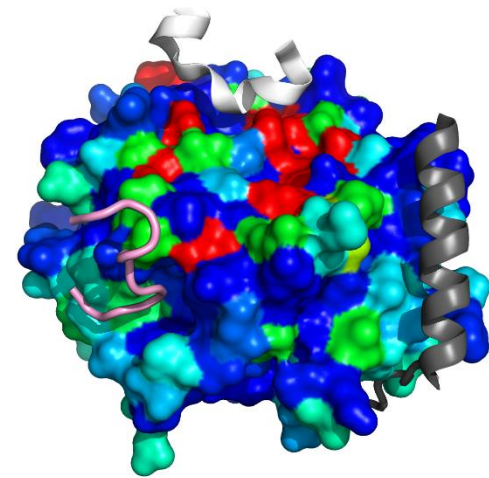
- Reported as novel fold...
- ... but in fact related to *Bacillus* toxin structures
- Both bind to host insect membranes
- Palmitate seen in Evf structure. Matches conserved region of toxins...
- **GESAMT** is an excellent CCP4 option



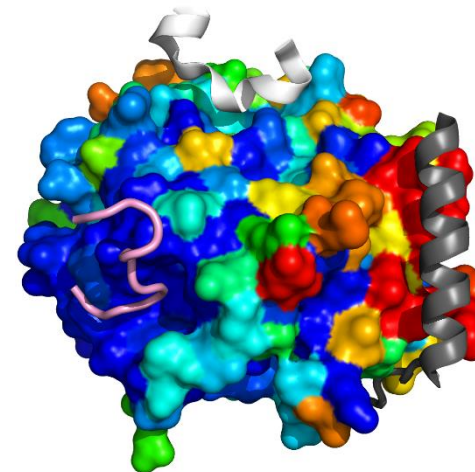
Some servers require thought...

- Consurf maps sequence conservation onto a structure revealing functional sites
- Excellent, general method, but results depend on sequence set chosen for mapping: selecting all or only near relatives gives different results. Either might be more appropriate for you

Mapping 300 homologues mixes different activities so no information on binding sites



But restricting to a single protein family shows only 'pink' site is function in both Diptera and Lepidoptera



...and finally, you're putting a
manuscript together

Calculating and presenting sequence alignments

Your sequence alignment

- Don't use ClustalW! It's 23 years old! Modern methods like MUSCLE, Probcons and MAFFT are much better

ClustalW misses relatively obvious RHG motif in some of diverse sequence set...

Sequence alignment from ClustalW. The alignment shows 10 sequences from various species. The RHG motif is circled in red in the original image. The alignment is as follows:

Sequence	150	160	170	180	190	200	210	220	230	
gi 548534 sp P00950 PMG1_YEAST/1-487	MPKLVLYRHGSEWNEK	----	NLFTGWV	DVKLSAKGQQ	----	EAARAGELLKEKKVY	PDV	----	LYTSKLSRAITANIALEKADR	
gi 3024420 sp P96121 GPMA_TREP/1-487	MKLVLIRHGESEWNL	----	NLFTGWT	DVPLTPRGES	----	EAQEGGRVLQEAQDFDL	----	----	CYTSYLRKRAIRTLNFVLQALDR	
gi 12751461 gb AAK07665.1 /1-487	-----	TRHGEKWNVE	----	RRMQGWQ	DSPLTEKGRQ	----	DAMRLCKRLEA	VELAA	----	IYTSYSGRALTAIEIVRGCR
gi 1169587 sp P32604 F26_YEAST/1-487	TS	PDYENSEPHVAADFLE	----	IRQYERF	YEPLDQKDK	----	DMTFIKLVNIEEVVINK	----	IRTVLESRIVYMMNIRPKPKY	
gi 1730554 sp P52086 COBC_ECOL/1-487	-----	MRLWLIHGESEWNL	----	GLYSCHA	PTPLTARGIE	----	QAQNLHTLLHG	VSF	----	VLCSELERAQITARLVLSDRQ
gi 3183165 sp P76502 SIXA_ECOL/1-487	-----	MQVFIMRHGDAALD	----	AASDS	VRPLTTNGCD	----	ERLMANWLKQKVEIER	----	VLVSPLRAEITLLEEVGDCLN	
gi 2895490 gb AAC38954.1 /1-487	LELPVDICIRHGTQGNTE	PRVFQGVQDYA	NNQLTQQCQQQAAAAATK	LEAMAAKEFI	PD	----	LLSSPLLRAVHTAQPFVDANK	----	----	
gi 15229917 ref NP_187168.1 /1-487	KLLPKRIILVRHGESEGNLDT	AAYTTTPDH	KIQLTDSGLLQAQEA	CARLHALISSNP	SSPEWRVYFV	VS	YDRTESTLREIGRSFSR	----	----	
gi 16130187 ref NP_416755.1 /1-487	-----	MLAFCRSLKSKY	IIILLALAAIAGL	THAAWSNGL	PR	----	INKTARLAQHPVVVLF	----	----	FERDORSTNQCLSDKT

... but MUSCLE gets it

Sequence alignment from MUSCLE. The alignment shows the same 10 sequences. The RHG motif is correctly identified and circled in red in the original image. The alignment is as follows:

Sequence	220	230	240	250	260	270	280	290	300		
gi 2895490 gb AAC38954.1 /1-537	-----	LELPVDICIRHGTQGNTE	PRVFQGVQDYAN	NNQLTQQCQQQAAAAATK	LEAMAAKEFI	PD	----	LLSSPLLRAVHTAQPFV	----		
gi 1730554 sp P52086 COBC_ECOLI/1-537	-----	MRLWLIHGESEWNL	GLYSCHA	PTPLTARGIE	QAQNLHTLLHGVSF	----	LV	LCSELERAQITARLV	----		
gi 12751461 gb AAK07665.1 /1-537	-----	TRHGEKWNVE	RRMQGWQ	DSPLTEKGRQ	DAMRLCKRLEA	VELAA	----	AI	----	YTSYSGRALTAIEIV	
gi 548534 sp P00950 PMG1_YEAST/1-537	-----	MPKLVLYRHGSEWNEK	NLFTG	----	WVDVKLSAKGQQ	EAARAGELLKEKKVY	----	PD	VL	----	YTSKLSRAITANIA
gi 3024420 sp P96121 GPMA_TREPA/1-537	-----	MKLVLIRHGESEWNL	NLFTG	----	WTDVPLTPRGES	EAQEGGRVLQEAQDF	----	FD	LC	----	YTSFLKRAIRTLNFV
gi 16130187 ref NP_416755.1 /1-537	-----	PVVVLFIRHAEFCDRSTNQCLSD	----	KTGITVKGTQDAREL	QNAFSADI	----	PDFDL	----	YSSNVRTI	SA	----
gi 3183165 sp P76502 SIXA_ECOLI/1-537	-----	MQVFIMRHGDAALDAASDSVR	----	PLTTNGCDESRLMANWLKQKVE	----	IE	RV	----	LVSPFLRAEITL	----	----
gi 15229917 ref NP_187168.1 /1-537	-----	KLLPKRIILVRHGESEGNLDTAAYTT	TPDH	KIQLTDSGLLQAQEA	CARLHALISSNP	SSPEWRVYFV	VS	YDRTESTLREIG	----	----	----
gi 1169587 sp P32604 F26_YEAST/1-537	----	YMMNIRPKPKYTWLSRHGESEYNVE	KKICG	----	DSSL	SEKGFYAKKLEQLVKESAGE	----	INLTV	----	WTSTLRTITAN	YL

Multiple methods in bioinformatics: Jalview.org

- All these alignment methods and more are available through Jalview on Dundee servers

Jalview 2.9.0b2

File Tools Vamsas Help Window

MAFFT Multiple Sequence Alignment of Retrieved from Uniprot

File Edit Select View Annotations Format Colour Calculate Web Service

Original Phosphorylation Site Predictions MAFFT Alignment Order

Alignment

Secondary Structure Prediction

Protein Disorder

Analysis

Conservation

Fetch DB References

<http://www.compbio.dundee.ac.uk/jabaws>

Tcofee with Defaults

Edit settings and run...

Run Tcofee with preset

Probcons with Defaults

Edit settings and run...

Muscle with Defaults

Edit settings and run...

Run Muscle with preset

MAfft with Defaults

Edit settings and run...

Run MAfft with preset

MSAProbs with Defaults

Edit settings and run...

GLProbs with Defaults

Edit settings and run...

Clustal

Realign with Clustal

ClustalO

Realign with ClustalO

Average distan...

File View

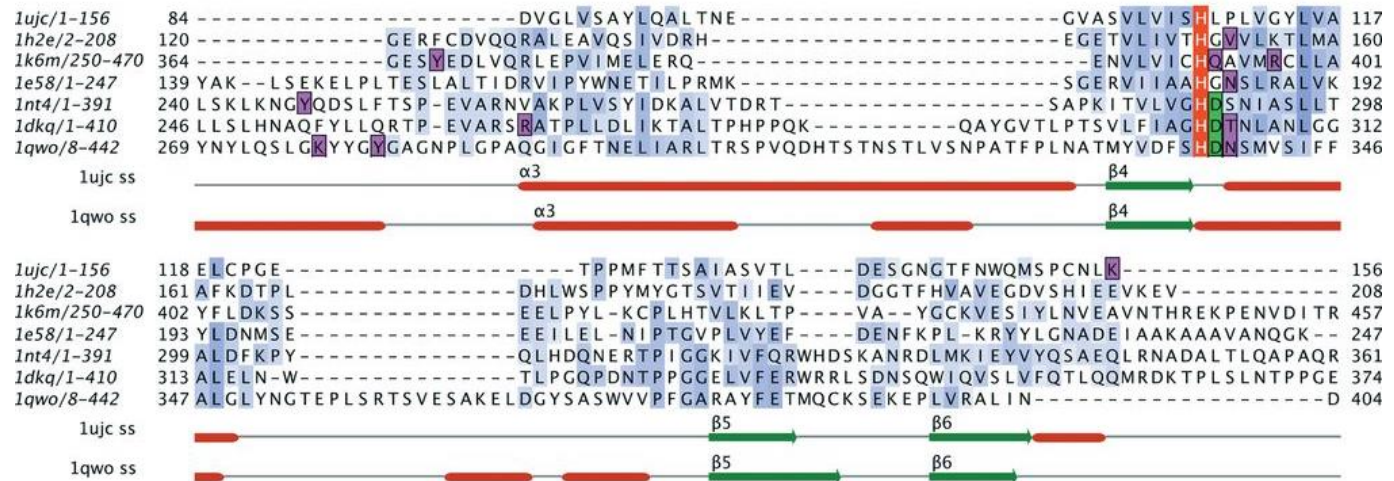
3D view for FER1_S...

Align with default settings

THR 89

Sequence 1 ID: FER_CAPAA Residue: PRO (36)

- Also helps you produce figures like this...



- ... rather than like this



Questions?

